

# Construction of Plasmid DNA for Heterologous Expression of Ester Forming Enzymes

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## Introduction

### Abstract

Developing a microbial system increasing production of sustainable solvents like ethyl lactate have environmental and economic impacts. Prior bioinformatics analysis showed ATF1, ATF2 and LgATF1 to be potential genes. In silico and in vitro laboratory studies were performed to confirm their functionality in ester biosynthesis. PCR was used to amplify both a small section of DNA (*cgATF1 gene*) and the plasmid (*pCRB204*) determined by primers. Then, CPEC cloning was performed where insert and vector get compatible ends to connect to each other. To finish, heat shock bacterial transformation in E.Coli of these genes was performed followed by Colony PCR and gel electrophoresis in order to verify that the desired gene is present in the plasmid.

### Motivation

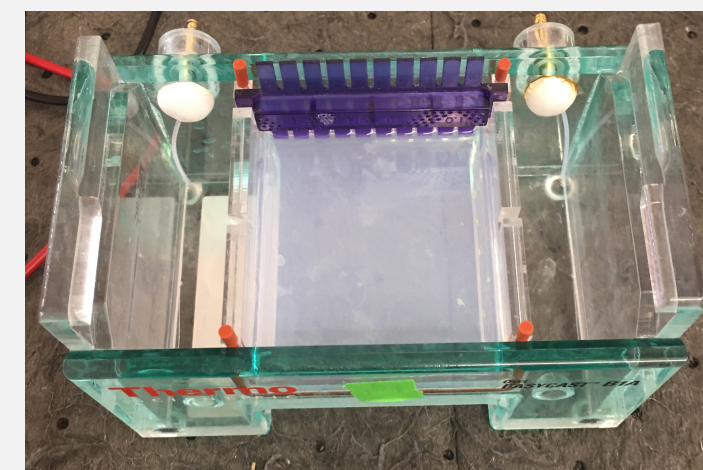
- Current chemical solvents' industries are petroleum-based, nonbiodegradable, toxic and non-renewable
- Ethyl lactate could be used as an biodegradable alternative, renewable
- Increasing the production of sustainable solvents such as ethyl lactate is urging
- Using agricultural products as raw materials
- Exploring more such genes that can improve its yield
- Exploring new esterase or acyl alcohol transferase genes
- The economic possibilities of its production as a green solvent is very consequent
- However the production means remains costly

Goal: Will ethyl lactate be able to be produced in a more efficient way?

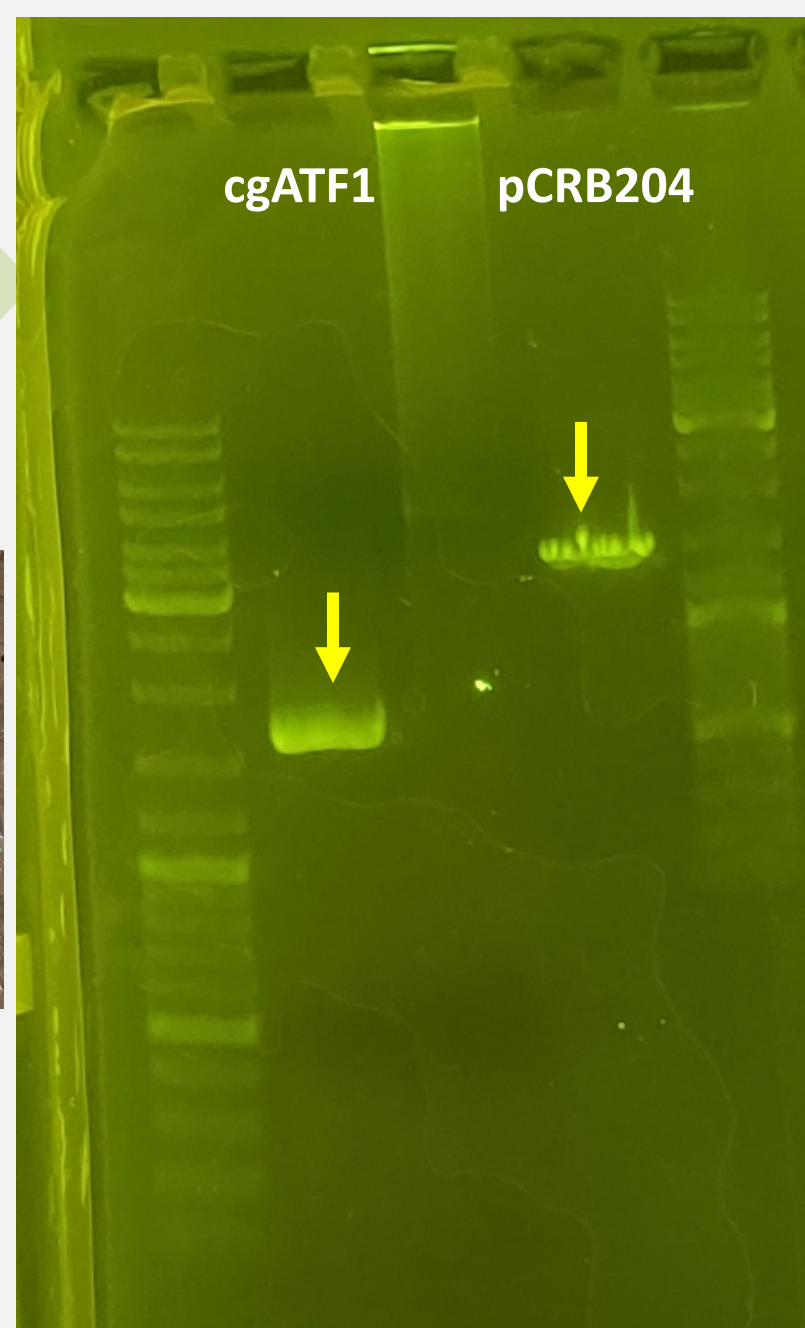
## Approach/Results/Discussion

### PCR Amplification of

Insert: *cgATF1 gene*  
Vector: *pCRB204*



Agarose gel electrophoresis



Gel purify the fragments

### CPEC Cloning

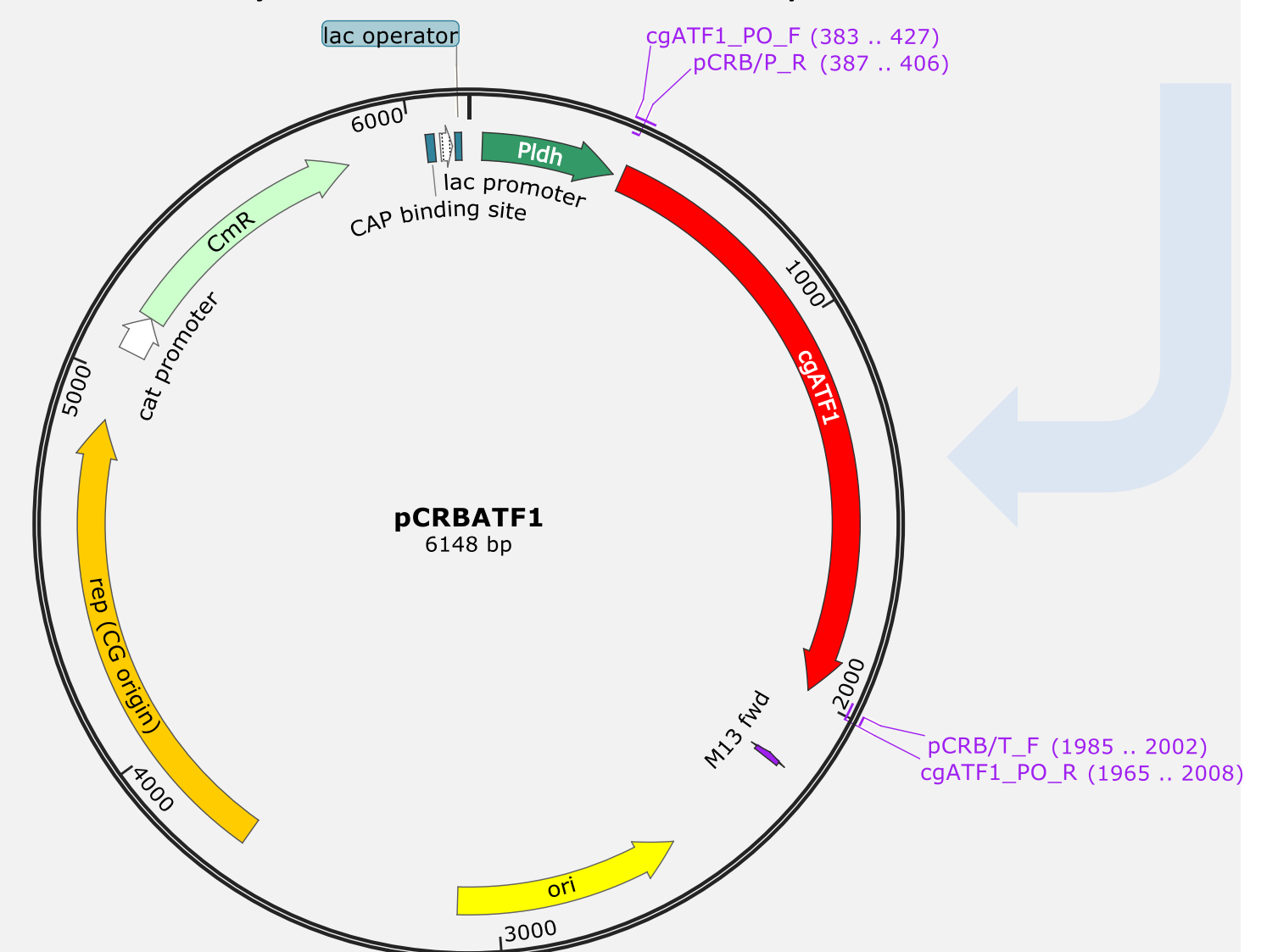
### CPEC REACTION MIX

Initial concentration	Volume per 20 µl reaction	Final amount per 20µl reaction
Phusion HF buffer (5x)	4 µl	1x
dNTP mix (40 nM)	0.4 µl	0.8nM
Phusion high-fidelity DNA polymerase (2 U µl <sup>-1</sup> )	0.2 µl	1 u
Vector DNA	Variable	100-200 ng
Insert gene library (-ies)	Variable	--
ddH <sub>2</sub> O	Up to 20 µl	

### CPEC REACTION PROTOCOL ON PCR MACHINE

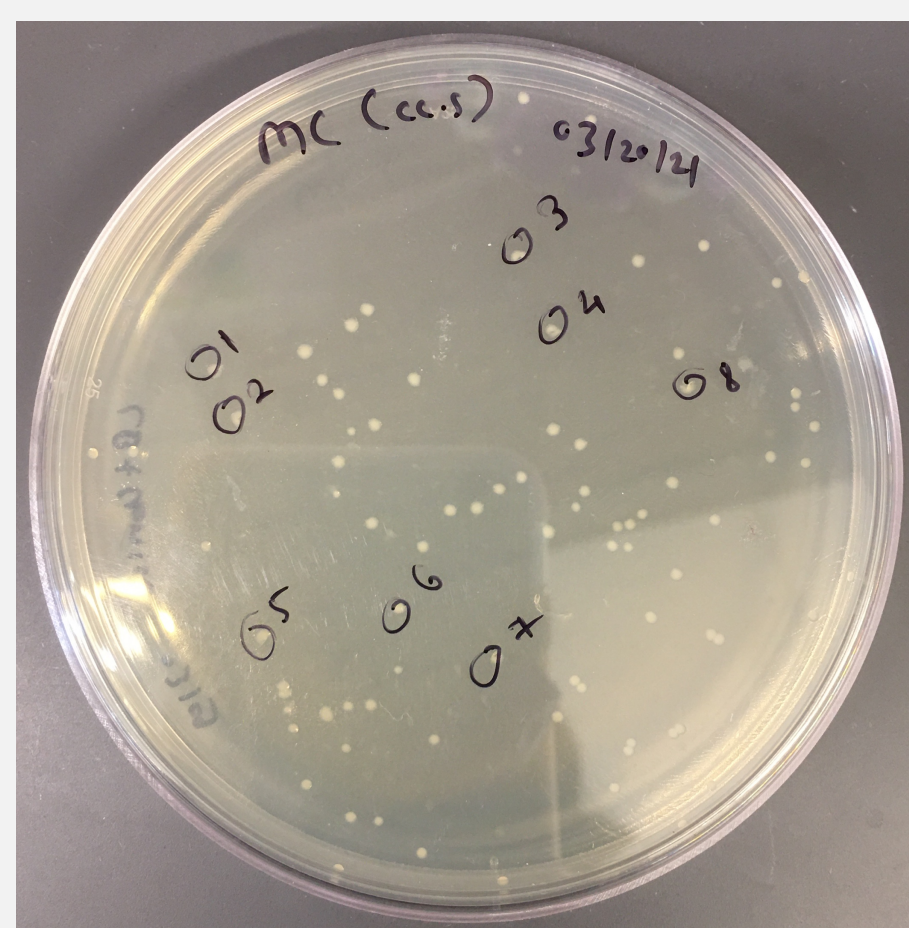
Cycle number	Denature	Slow ramp anneal	Anneal	Extend
1	98°C, 30s	---	---	---
2-26	98°C, 10s	70 to 55°C, 3min	55°C, 30s	72°C, 10-20s per kb
27	---	---	---	72°C, 5 min

### Plasmid synthesized due to overlaps in CPEC

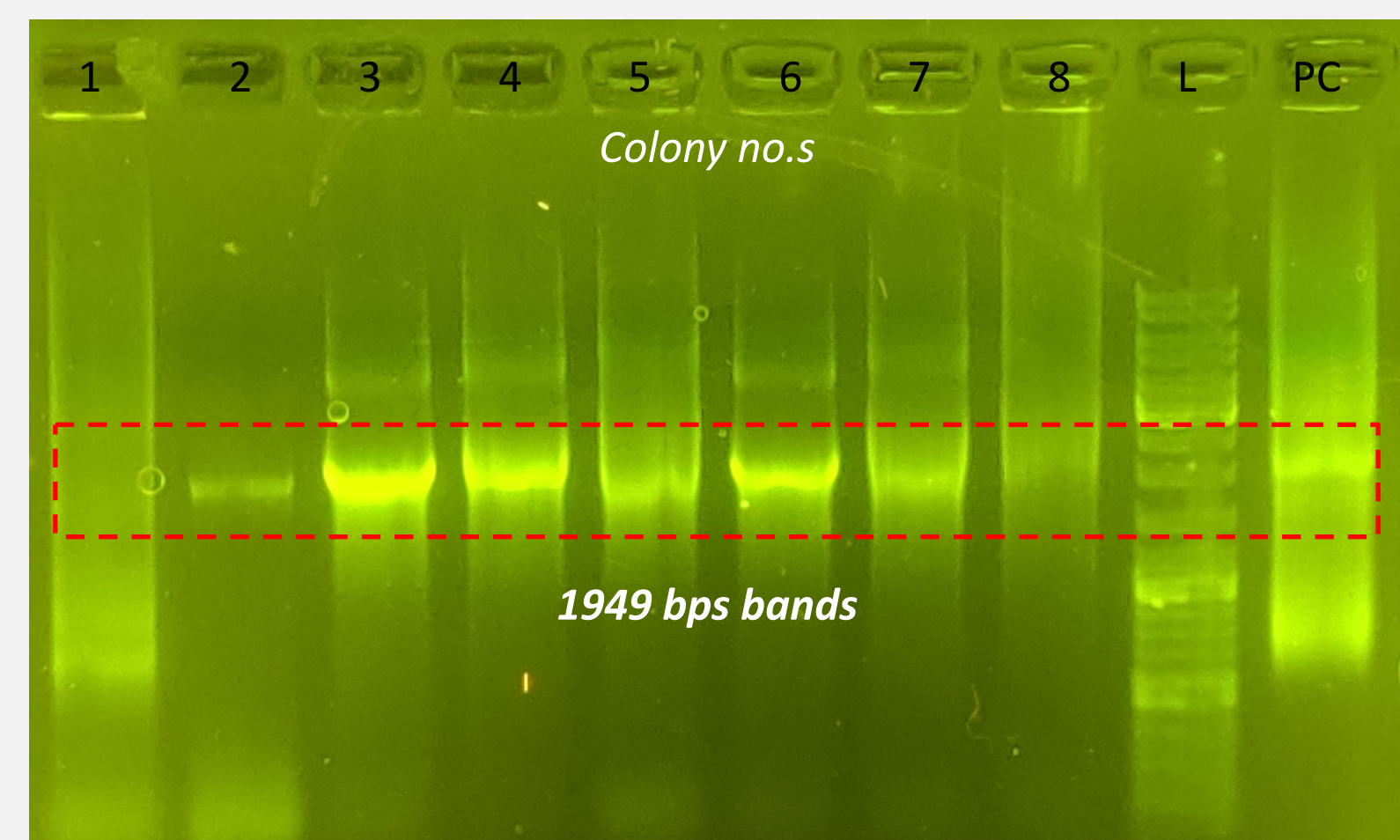


### Transformation In E.Coli

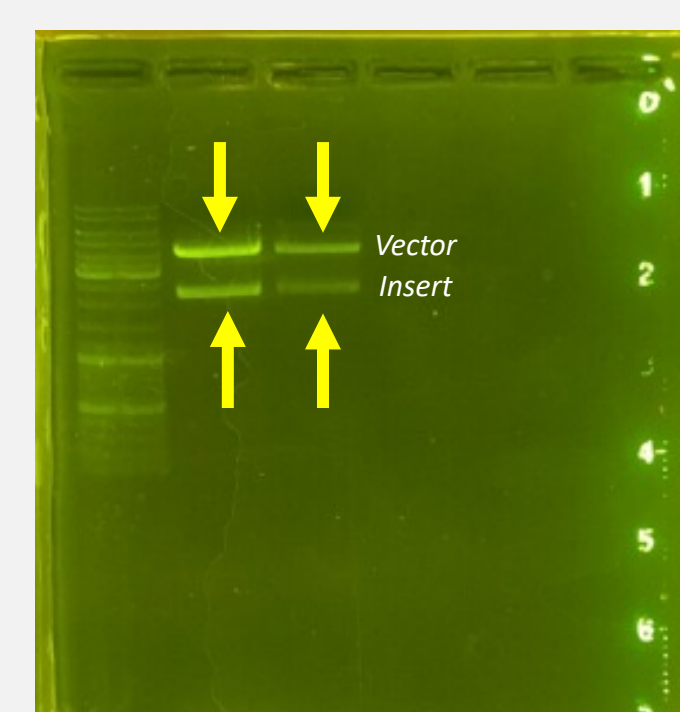
### Transformant colonies



### Colony PCR



### Successful cloning



### Inoculate the positive colonies

### Acknowledgements:

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- Apurv Mhatre

### Future prospect:

Perform plasmid-prep and purify the plasmid for transforming into *Corynebacterium glutamicum*